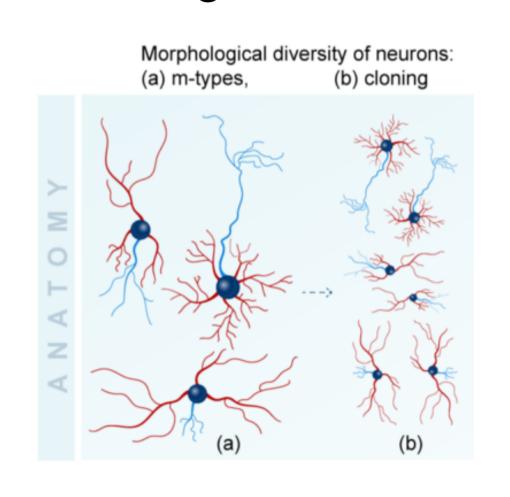
RT

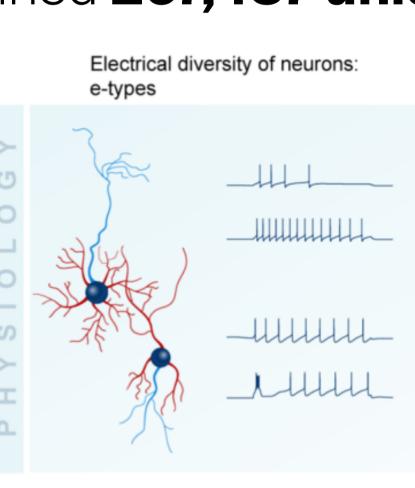


<sup>2</sup>Department. of Psychiatry, <sup>3</sup>Dept. of Physiology, University of Toronto, Canada 4Krembil Ctr. for Neuroinformatics, Center for Addiction and Mental Health, Toronto, Canada

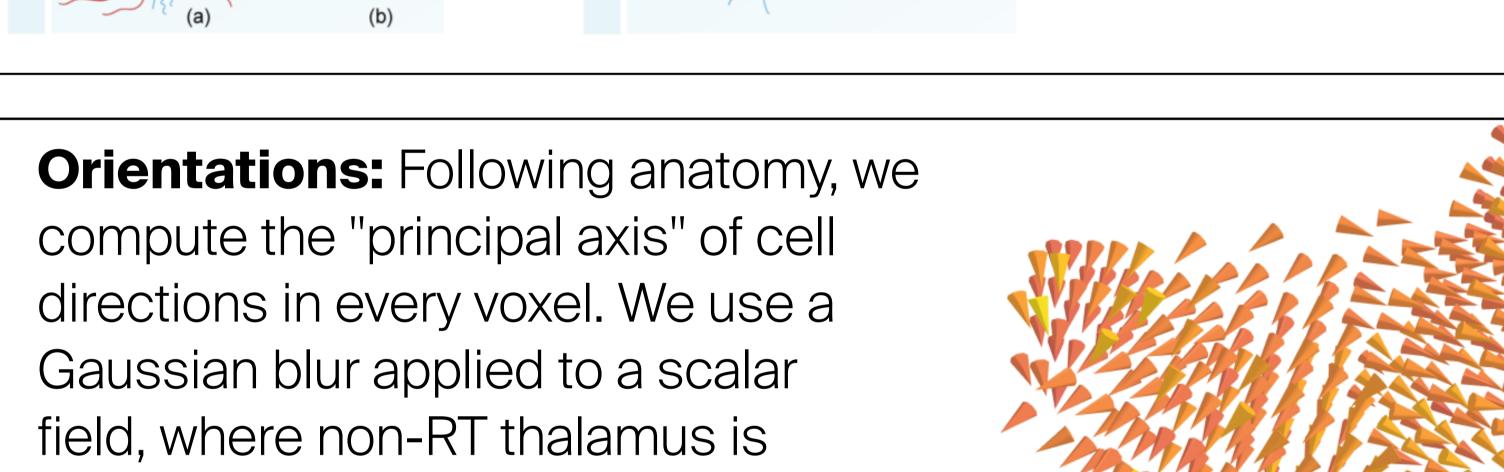
## Building an Atlas-based PO, RT, and VPL Circuit

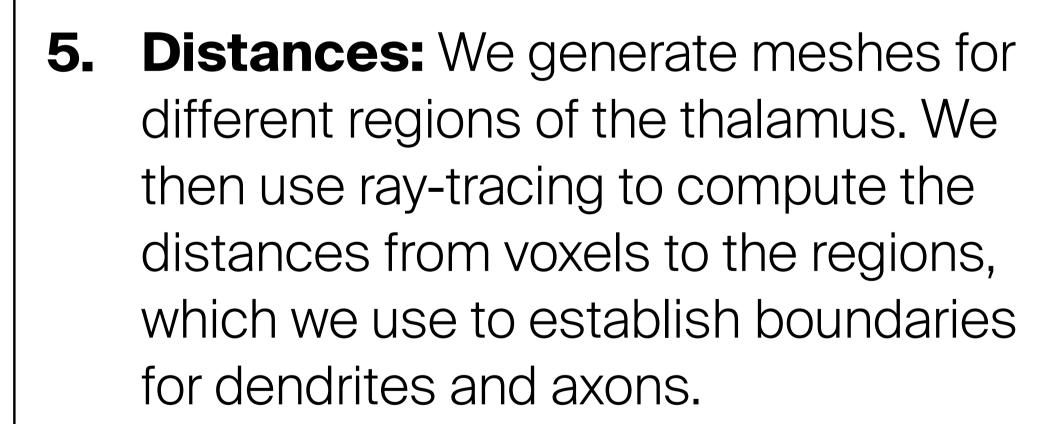
- Morphology: First, we obtain reconstructions of individual neurons from experimental data. We then "clone" these cells, varying and swapping neurites for each cell-type. For the thalamus, we use 3 classes of M-type: TC, IN, and RT. (lavarone et al., 2023)
- Electrophysiology: Next, using ion channel parameters optimized against experimental data, we characterize different classes of neurons. These electrophysiological **E-types** are then fit to their relevant morphologies. For the thalamus, we use 5 E-type classes and organize our celltypes into 5 unique ME-type classes. We have generated a combined 267,487 unique thalamus cells.



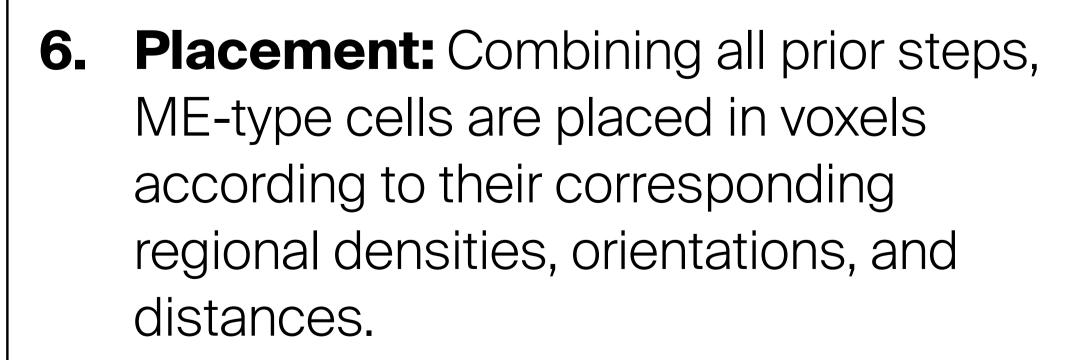




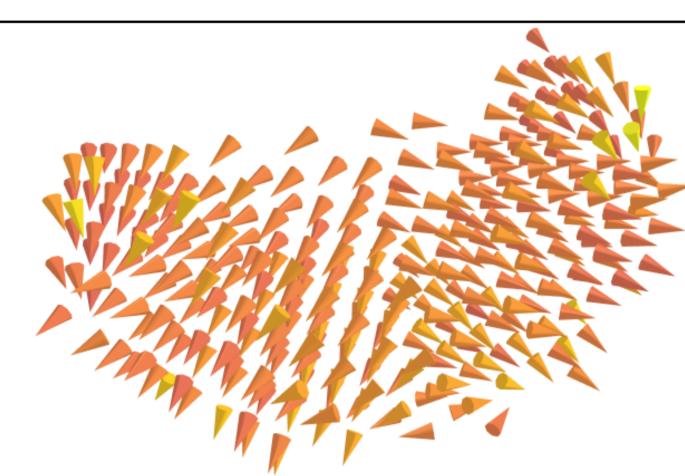




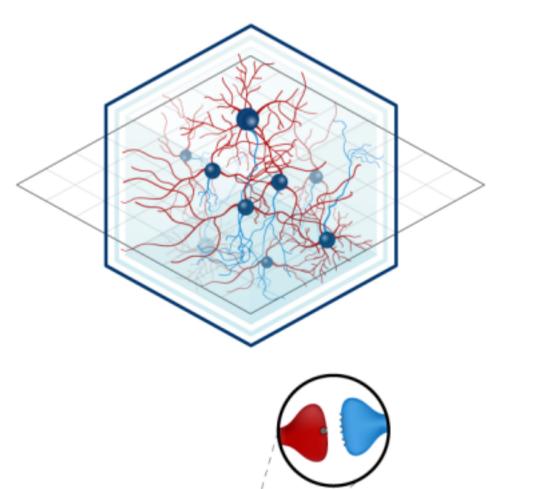
the source and RT is the target.

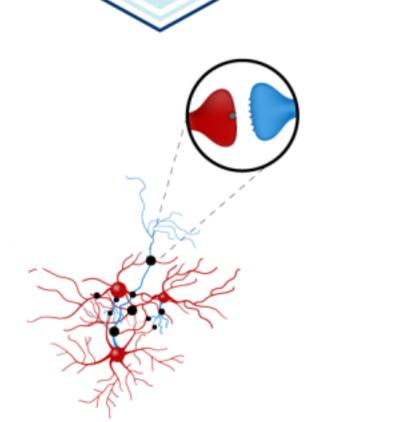


7. Synapses: Potential synapses are identified at relevant appositions ("touches") between celltypes. Synapses are then "pruned" to appropriate levels (Reimann et al., 2015).

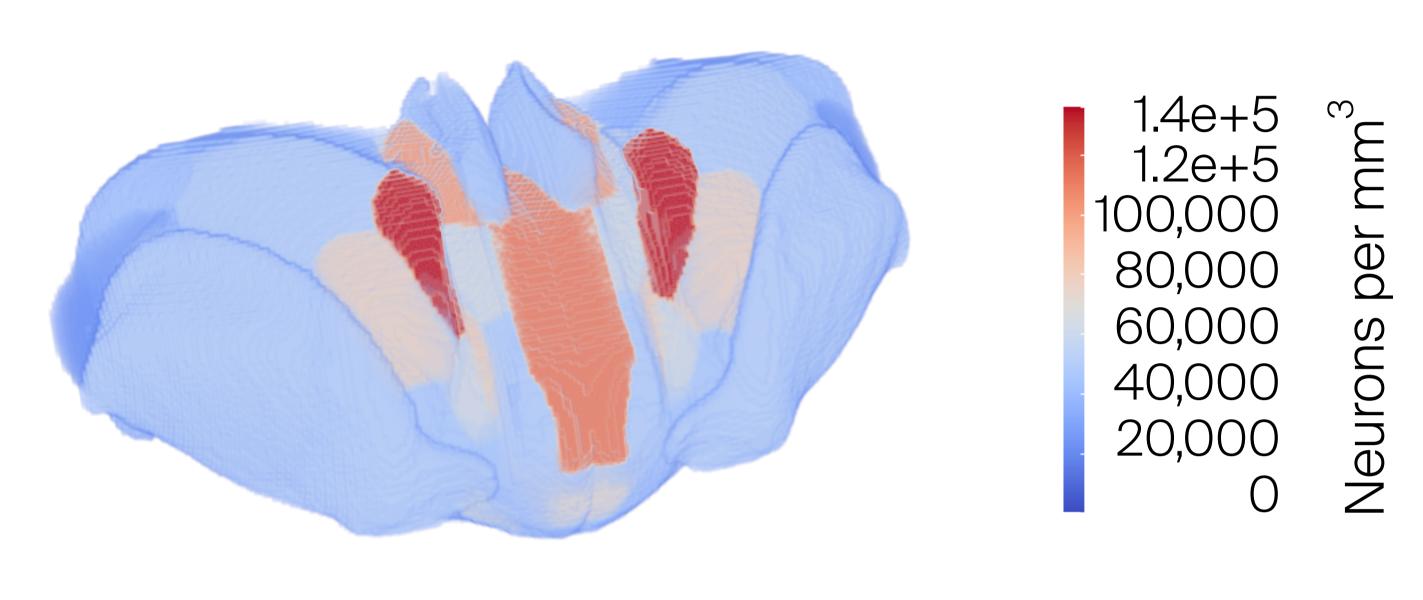




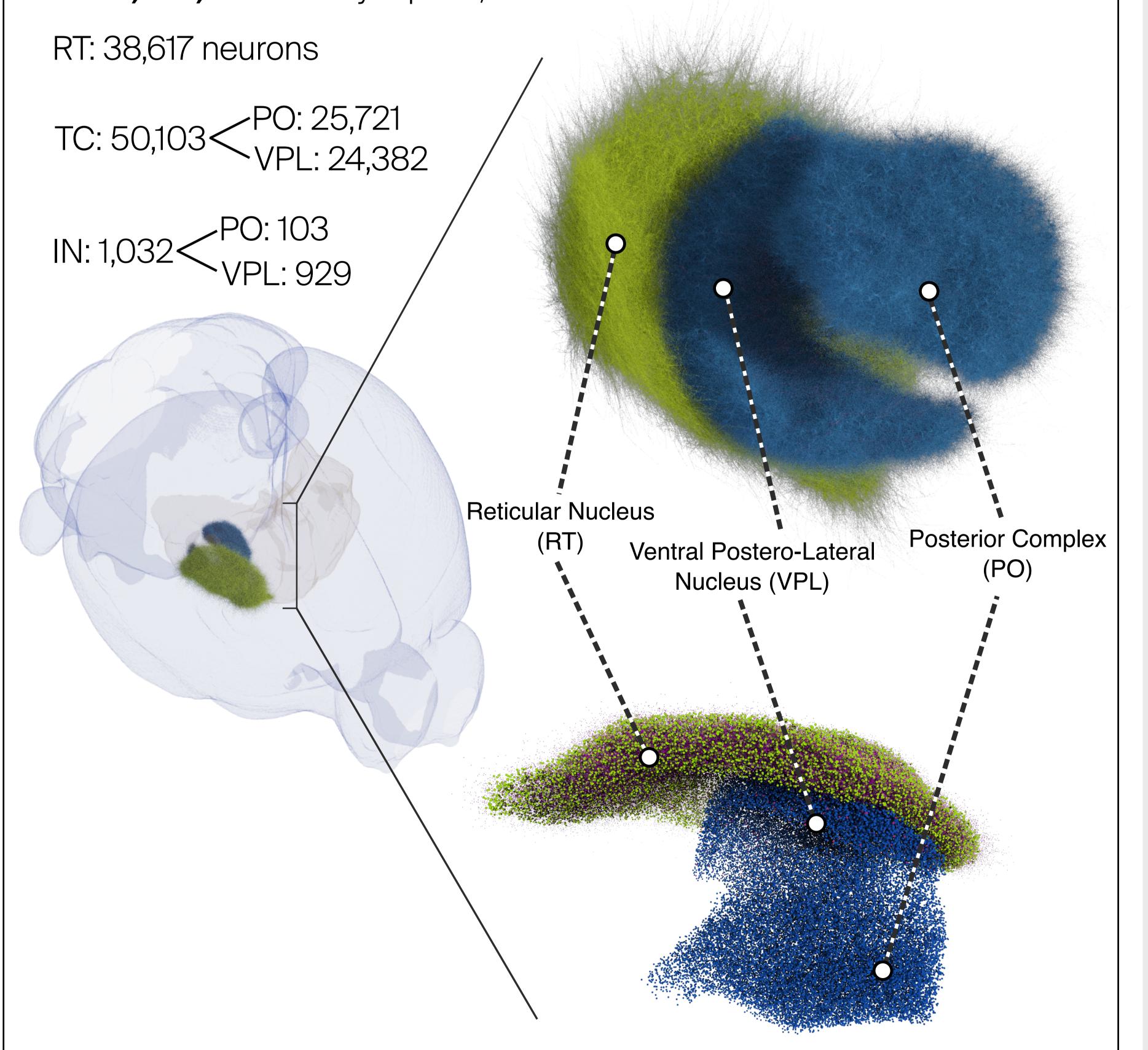




**Densities:** Using the spatial mapping of the Allen Mouse Brain Atlas (CCFv3) (Wang et al., 2020) and volumetric neuron densities from the Blue Brain Cell Atlas (Rodarie et al., 2022) and literature, we assign our cells to different thalamic regions in space.



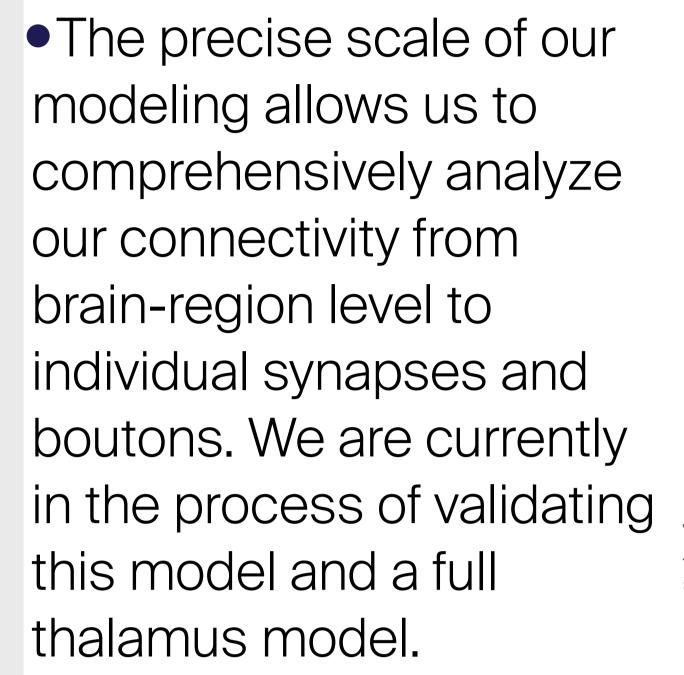
8. Final Circuit: The circuit ultimately comprises 89,752 total cells, **76,351,201** total synapses, and **3.576** mm<sup>3</sup> volume.

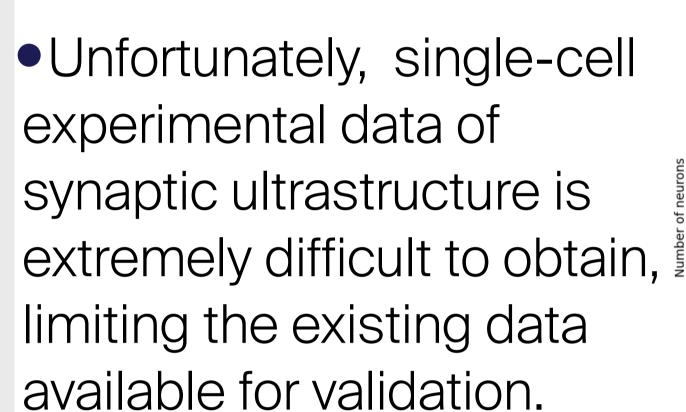


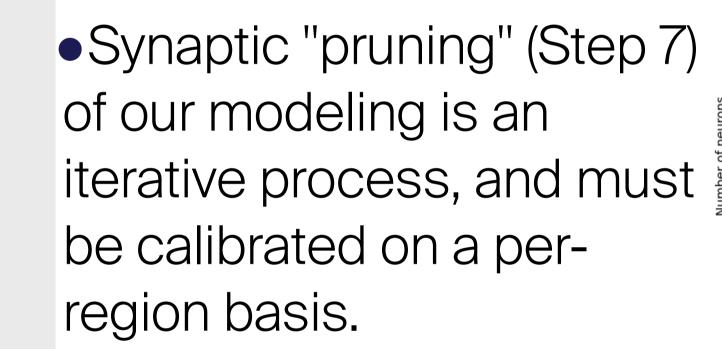
## Connectivity Validations

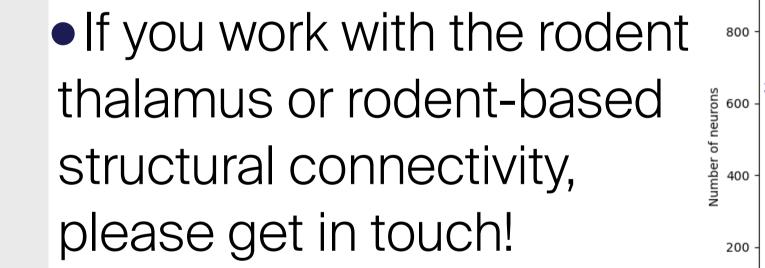
TC

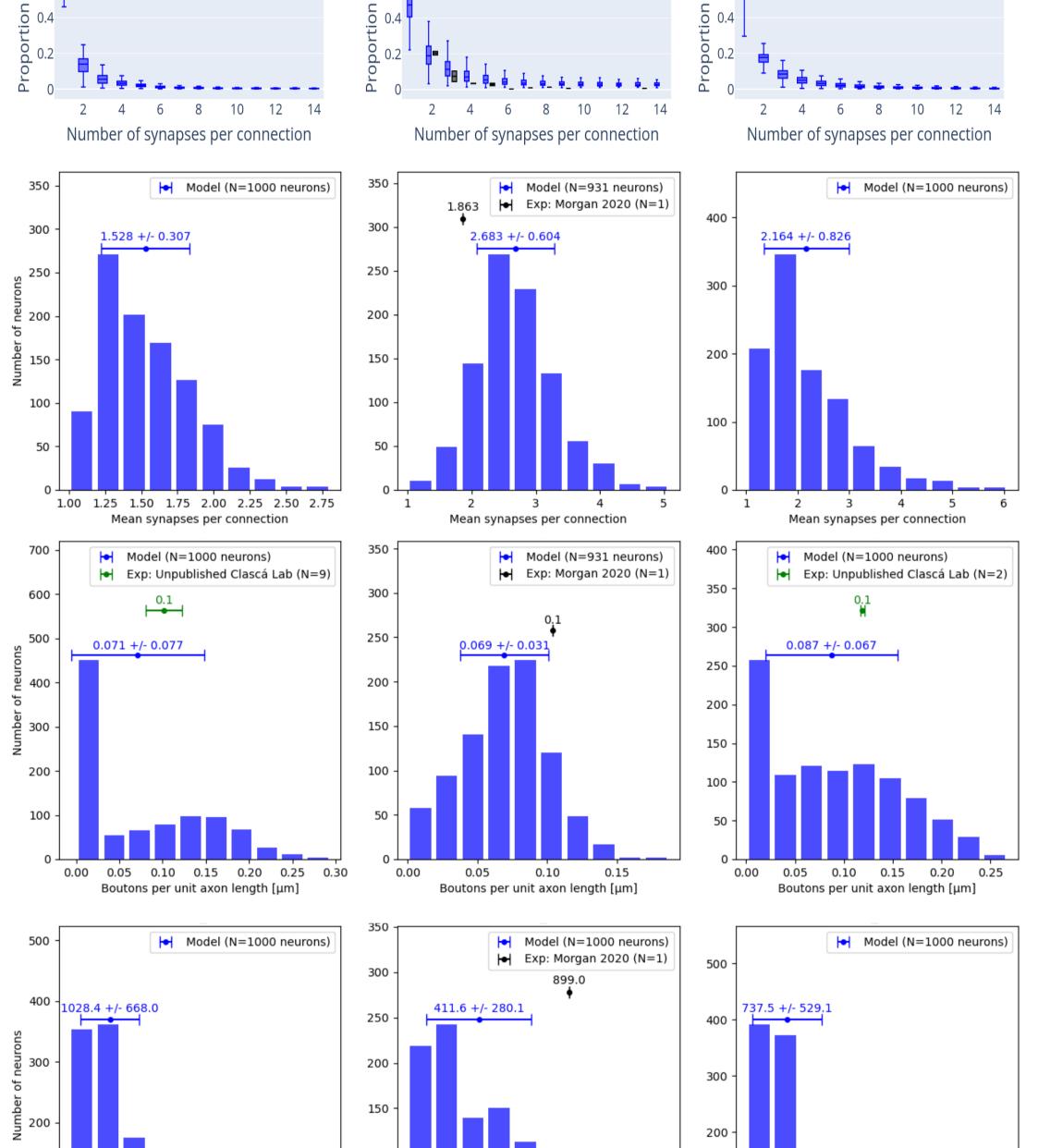
<sup>1</sup>Blue Brain Project, École polytechnique fédérale de Lausanne (EPFL), Campus Biotech, Genève, Switzerland











H Exp: Morgan 2020 (N=1)

IN

■ Exp: Morgan 2020 (N=1)

## References and Acknowledgements

lavarone E, Simko J, Shi Y, et al. Thalamic control of sensory processing and spindles in a biophysical somatosensory thalamoreticular circuit model of wakefulness and sleep. Cell Reports. 2023;42(3):112200. doi:10.1016/j.celrep.2023.112200 . Rodarie D, Verasztó C, Roussel Y, et al. A method to estimate the cellular composition of the mouse brain from heterogeneous datasets. PLOS Computational

. Wang Q, Ding SL, Li Y, et al. The Allen Mouse Brain Common Coordinate Framework: A 3D Reference Atlas. Cell. 2020;181(4):936-953.e20.

4. Morgan JL, Lichtman JW. An Individual Interneuron Participates in Many Kinds of Inhibition and Innervates Much of the Mouse Visual Thalamus. Neuron. . Reimann MW, King JG, Muller EB, Ramaswamy S, Markram H. An algorithm to predict the connectome of neural microcircuits. Frontiers in Computational Neuroscience. 2015;9. Accessed February 3, 2023. https://www.frontiersin.org/articles/10.3389/fncom.2015.00120

6. Meyer HS, Egger R, Guest JM, Foerster R, Reissl S, Oberlaender M. Cellular organization of cortical barrel columns is whisker-specific. Proceedings of the National Academy of Sciences. 2013;110(47):19113-19118. doi:10.1073/pnas.1312691110 7. Pinault D, Deschênes M. Projection and innervation patterns of individual thalamic reticular axons in the thalamus of the adult rat: a three-dimensional, graphic, and morphometric analysis. J Comp Neurol. 1998;391(2):180-203. doi:10.1002/(sici)1096-9861(19980209)391:2<180::aid-cne3>3.0.co;2-z 8. Markram H, Muller E, Ramaswamy S, et al. Reconstruction and Simulation of Neocortical Microcircuitry. Cell. 2015;163(2):456-492. doi:10.1016/j.cell.2015.09.029

Thank you to Elvis Boci and Cyrille Favreau for visualization help! This study was supported by funding to the Blue Brain Project, a research center of the École polytechnique fédérale de Lausanne (EPFL), from the Swiss government's ETH Board of the Swiss Federal Institutes of Technology.

► Exp: Pinault 1998 (N=1